

Epi proColon[®]

Epi proColon Plasma Quick Kit (M5-02-001)

Epi proColon Sensitive PCR Kit (M5-02-002)

Epi proColon Control Kit (M5-02-003)

For *in vitro* diagnostic use only

Read and follow all instructions for use prior to using this test.



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M5-02-001, M5-02-002, M5-02-003



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1. Name and Intended Use

Intended Use Statement

The Epi proColon test is a qualitative *in vitro* diagnostic test for the detection of methylated Septin9 DNA in EDTA plasma derived from patient whole blood specimens. Methylation of the target DNA sequence in the promoter region of the *SEPT9_v2* transcript has been associated with the occurrence of colorectal cancer (CRC)¹. The test uses a real-time polymerase chain reaction (PCR) with a fluorescent hydrolysis probe for the methylation specific detection of the Septin9 DNA target.

The test is indicated to screen patients for colorectal cancer who are defined as average risk for colorectal cancer (CRC) by current CRC screening guidelines. Patients with a positive Epi proColon test result should be referred for diagnostic colonoscopy. Men and women 50 to 85 years of age were included in the Epi proColon clinical trial. The Epi proColon test results, together with the physician's assessment of history, other risk factors, and professional guidelines, may be used to guide patient management.

The Epi proColon test is for use with the Applied Biosystems 7500 Fast Dx Real-Time PCR Instrument.

Warnings

- The Epi proColon test is not intended to replace colorectal screening by colonoscopy.
- The Epi proColon test is not intended to screen persons under the age 50 who are considered to be at average risk for colorectal cancer.
- Positive Epi proColon test results are not confirmatory evidence for the presence of colorectal cancer. Patients with a positive Epi proColon test result should be referred for diagnostic colonoscopy.
- A negative Epi proColon test result does not guarantee absence of cancer. Patients with a negative Epi proColon test result should be advised to continue participating in a colorectal cancer screening program that also includes colonoscopy, fecal tests and/or other recommended screening methods.
- Positive test results have been observed in clinically diagnosed patients with chronic gastritis, lung cancer and in pregnant women^{6,7}.

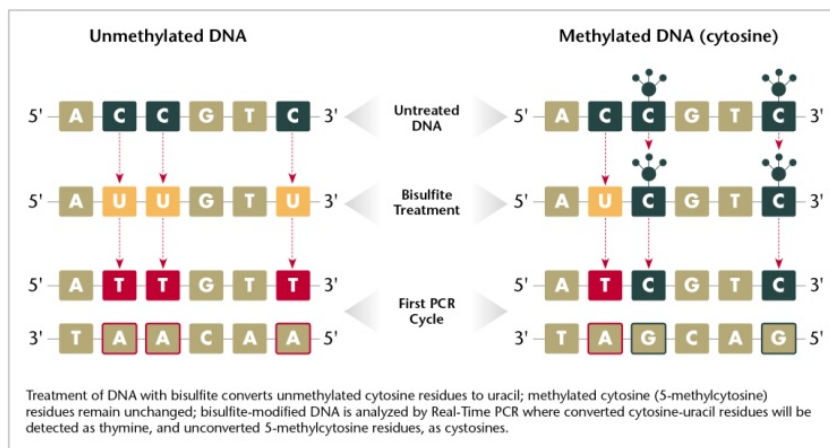
2. Summary and Explanation

The Epi proColon test is an *in vitro* polymerase chain reaction (PCR) assay for the qualitative detection of methylated Septin9 DNA isolated from 3.5 mL of patient plasma. Cytosine residues in the v2 region of the Septin9 gene become methylated in colorectal cancer (CRC) tissue but not in normal colon mucosa. This aberrant methylation can be detected by specific amplification of Septin9 DNA present in the plasma sample. Detection of CRC DNA in plasma using the methylated Septin9 DNA biomarker has been demonstrated in multiple case control studies of CRC patients and colonoscopy-verified negative controls¹⁻⁴, as well as in two multicenter clinical evaluations. The blood-based Epi proColon test offers patients who are unwilling, unable or do not undergo screening by other recommended methods an alternative option to participate in a CRC screening program.

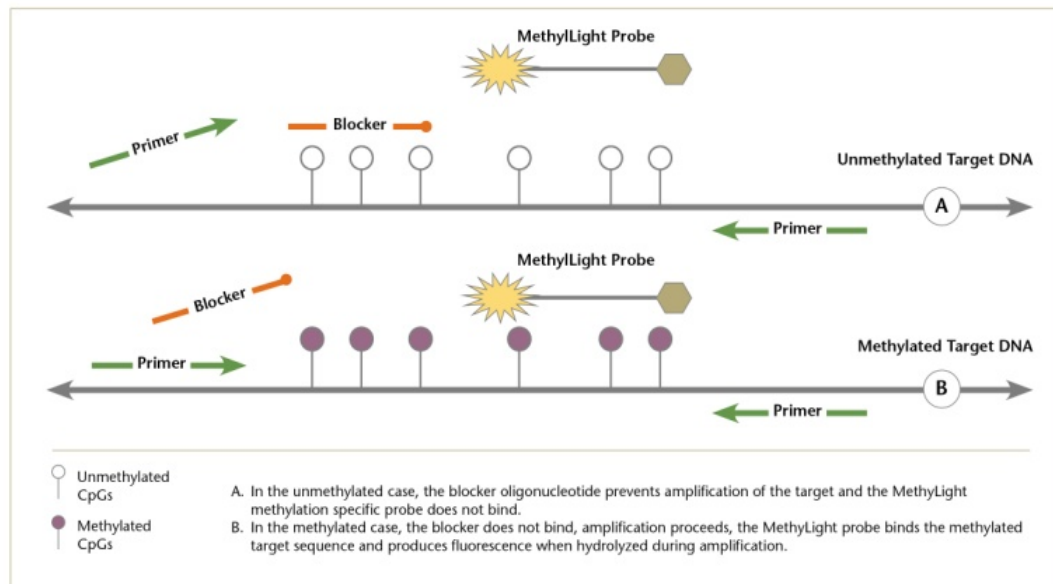
3. Principles of the Procedure

The Epi proColon test involves two primary procedural phases:

First, free DNA is extracted from plasma and treated with bisulfite using the Epi proColon Plasma Quick Kit (M5-02-001). DNA is extracted from patient plasma by binding to magnetic beads. Impurities are removed from the magnetic beads by a wash step. The purified DNA is then released from the beads by suspension in an elution buffer, and treated with bisulfite reagents to produce bisulfite converted DNA (bisDNA). The bisDNA is then repurified using magnetic beads. Bisulfite conversion is the method of choice for analyzing DNA methylation. It is based on the nucleophilic addition of a bisulfite ion to a cytosine nucleotide and a subsequent deamination reaction to yield uracil sulfonate; 5-methylcytosine (methylated cytosine) is protected from deamination by the methyl group. Thus, the sequence of bisDNA differs from the input DNA by the substitution of uracil nucleotides for unmethylated cytosines, while retaining methyl cytosine bases.



Second, bisDNA is assayed by a duplexed real time PCR (M5-02-002) with simultaneous detection of the target methylated Septin9 DNA and the internal control ACTB (β -actin) DNA. The control is designed to establish that sufficient input DNA is included in the reaction. Positive and Negative Controls are provided with the Epi proColon Control Kit (M5-02-003) and are required for each test run.



The duplex Epi proColon test detects a bisDNA sequence containing methylated CpG sites within the v2 region of the *SEPT9* gene and a bisDNA sequence from the *ACTB* gene with no methylated CpG sites. The Septin9 assay discriminates methylated cytosines from cytosines using a combination of a blocker oligonucleotide and a methylation specific probe. The reaction comprises primers that are placed in regions lacking CpG dinucleotides, a blocker specific for bisulfite converted unmethylated sequences within the region is added to preferentially amplify methylated sequences, and a methylated Septin9 specific fluorescent detection probe to exclusively identify methylated sequences amplified during the duplex PCR reaction⁵.

4. Precautions and Safety

Patient Precautions

- The Epi proColon test is an alternative screening method for patients who are defined as average risk for colorectal cancer by current screening guidelines, and who are unwilling, unable or do not undergo screening by other recommended screening methods.
- The Epi proColon test has not been evaluated in persons:
 - Considered to be at higher risk for developing colorectal cancer, or with a previous history of colorectal polyps or colorectal cancer. Persons at higher risk include those with a family history of colorectal cancer, particularly with two or more first-degree relatives with colorectal cancer, or one or more first degree relative(s) less than 50 years of age with colorectal cancer.
 - With known hereditary non-polyposis colorectal cancer (HNPCC) or familial adenomatous polyposis (FAP).
 - With anorectal bleeding, hematochezia, or with known iron deficiency anemia.
- Detection of colorectal cancer is dependent on the amount of free circulating tumor DNA in the specimen and may be affected by sample collection methods, sample storage, patient factors and tumor stage.
- There is insufficient evidence to report programmatic sensitivity of the Epi proColon test over an established period of time.

- CRC Screening guideline recommendations vary for persons over the age of 75. The decision to screen persons over the age of 75 should be made on an individualized basis in consultation with a healthcare provider.
- The Epi proColon test demonstrated non-inferiority to a FIT test (OC FIT-CHEK® Polymedco, Inc.), for sensitivity but not for specificity, indicating that the Epi proColon test exhibited a higher rate of false positive results compared to the FIT test. See Performance Characteristics in Section 13.
- Test results should be interpreted by a healthcare professional.

Laboratory Precautions Related to Real-Time PCR

- The Epi proColon test is for *in vitro* diagnostic use only
- This procedure is for professional laboratory use only and assumes familiarity with DNA extraction methods and real time PCR assays
- Compliance with good laboratory practices is essential to minimize the risk of cross-contamination between samples during and after the DNA extraction, bisulfite conversion, and purification procedure
- Use only single-use pipettes and filter tips to prevent cross-contamination of the patient sample
- Use of reference pipettes for pipetting extracted and bisulfite treated DNA is strongly recommended
- Good technique is important to prevent the introduction of nucleases into samples during the extraction procedure
- Do not freeze extracted DNA
- Epi proColon bisulfite solution is sensitive to oxygen contact; use only unopened tubes of Epi proColon Bisulfite Solution; do not store but discard any left-over solution
- When removing liquid from microtubes in multiple steps in the procedure, take care not to remove magnetic beads
- Strict separation of pre-PCR activities (e.g., plasma DNA extraction and purification, PCR setup) and post-PCR activities (e.g., real-time PCR) is highly recommended to prevent contamination by amplicons generated from previous PCR testing
- To prevent the release of any PCR product, used PCR plates should be placed in a resealable plastic bag immediately after removal from the PCR instrument, and the bag closed and disposed of in a dedicated PCR waste container
- Never open a used PCR plate or store a used PCR plate outside of the PCR instrument

Additional Precautions

- Do not mix kit components between kit lots
- Do not use kits or kit components beyond their stated expiration date
- Do not freeze whole blood K₂EDTA blood or blood tubes
- The Epi proColon test kits do not contain infectious substances or agents that may cause disease in humans or animals
- All patient blood and plasma specimens should be handled as though they are capable of transmitting disease. Observe universal precautions and safe laboratory procedures as specified in the OSHA Standard on Bloodborne Pathogens, CLSI Document M29-A3, and any other appropriate biosafety practices as required by your laboratory.
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5. Materials Provided

The Epi proColon test is comprised of the Epi proColon Plasma Quick Kit (M5-02-001), the Epi proColon Sensitive PCR Kit (M5-02-002) and the Epi proColon Control Kit (M5-02-003).

Contents

Table 1: Contents of the Epi proColon Plasma Quick Kit (M5-02-001)

Reagent	Containers	Volume	Storage Temperature
Epi proColon Lysis Binding Buffer	1 bottle	125 mL	15°C to 30°C
Epi proColon Wash A Concentrate	1 bottle	60 mL	15°C to 30°C
Epi proColon Magnetic Beads	1 bottle	4 mL	15°C to 30°C
Epi proColon Wash B Concentrate	1 bottle	7 mL	15°C to 30°C
Epi proColon Elution Buffer	1 tube	6 mL	15°C to 30°C
Epi proColon Bisulfite Solution	4 tubes	1.9 mL each	15°C to 30°C
Epi proColon Protection Buffer	1 tube	1 mL	15°C to 30°C

Table 2: Contents of the Epi proColon Sensitive PCR Kit (M5-02-002)

Reagent	Containers	Volume	Storage Temperature
Epi proColon PCR Mix	2 tubes	810 µl each	-25°C to -15°C
Epi proColon Polymerase	1 tube	85 µl	-25°C to -15°C

Table 3: Contents of the Epi proColon Control Kit (M5-02-003)

Reagent	Containers	Volume	Storage Temperature
Epi proColon Negative Control	6 tubes	3.65 mL each	-25°C to -15°C
Epi proColon Positive Control	6 tubes	3.65 mL each	-25°C to -15°C

Safety Information

When working with chemicals, always wear a laboratory coat and disposable gloves. Clean contaminated surfaces with water. For more information, please consult the respective material safety data sheets (MSDS) available on our webpage (tbd).

Epi proColon Lysis Binding Buffer and Epi proColon Wash A Concentrate:

contain TRITON X-100 and Guanidinium thiocyanate

Hazard statements: EUH032: Contact with acids liberates very toxic gas; H302: Harmful if swallowed; H312: Harmful in contact with skin; H318: Causes serious eye damage; H332: Harmful if inhaled; H412: Harmful to aquatic life with long lasting effects.

Precautionary statements: P261: Avoid breathing aerosols; P273: Avoid release to the environment; P301 + P312: IF SWALLOWED: Call a POISON CENTER or doctor/physician if you feel unwell; P302 + P352: IF ON SKIN: Wash with plenty of soap and water; P305 + P351 + P338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.



DANGER

Epi proColon Bisulfite Solution: contains aqueous solution of Ammonium bisulfite (Ammonium hydrogen sulfite)

Hazard statements: EUH031: Contact with acids liberates toxic gas; H314: Causes severe skin burns and eye damage.

Precautionary statements: P264: Wash hands thoroughly after handling; P271: Use only outdoors or in well-ventilated areas; P280: Wear protective gloves/protective clothing; P305+351+338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing; P312: Call a POISON CENTER or doctor/physician if you feel unwell.



DANGER

Epi proColon Protection Buffer: contains 6-hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid, Tetrahydrofurfuryl alcohol

Hazard statements: H319: Causes serious eye irritation.

Precautionary statements: P305 + P351 + P338: IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.



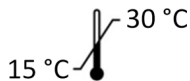
WARNING

Epi proColon Wash B Concentrate, Epi proColon Elution Buffer, Epi proColon Magnetic Beads, Epi proColon PCR Mix, Epi proColon Polymerase, Epi proColon Positive Control, and Epi proColon Negative Control are not harmful.

6. Storage and Stability

Reagents provided with the Epi proColon Plasma Quick Kit (M5-02-001), Epi proColon Sensitive PCR Kit (M5-02-002) and Epi proColon Control Kit (M5-02-003) are stable until the expiration date when stored and handled as directed. Do not use material past expiration date. Do not mix components from different kit lots.

Epi proColon Plasma Quick Kit (M5-02-001)



Store all reagents of the Epi proColon Plasma Quick Kit at 15 to 30 °C.

Epi proColon Bisulfite Solution is sensitive to oxygen contact. Use only unopened tubes of Epi proColon Bisulfite Solution. Discard used tubes!

Store reconstituted Epi proColon Wash A Buffer and reconstituted Epi proColon Wash B Buffer at 15 to 30 °C for up to 6 weeks.

After first use store all reagents at 15 to 30 °C for up to 6 weeks.

Epi proColon Sensitive PCR Kit (M5-02-002)



Store Epi proColon PCR Mix and Epi proColon Polymerase at -25 to -15 °C.

Each Epi proColon PCR Mix tube may be thawed and refrozen one time. After first use store all reagents at -25 to -15 °C for up to 6 weeks.

Epi proColon Control Kit (M5-02-003)



Store Epi proColon Control Kit at -25 to -15 °C.

7. Materials Required But Not Provided

The following general laboratory equipment is required to perform the Epi proColon test. All laboratory equipment should be installed, calibrated, operated, and maintained according to the manufacturer's recommendations.

Following, tables for required and special equipment and consumables are provided along with suggested vendors and catalog and part numbers.

General Laboratory Equipment

Required Equipment	Suggested Equipment
Tube racks for 15 mL tubes and 2 mL tubes	VWR International, LLC, catalog no. 82010-752, or equivalent
Rotator with adjustable angle	GLAS-COL 099A RD4512, VWR International, catalog no. 62404-006, or equivalent
Vortex Mixer	VWR International, LLC, catalog no. 58816-121, or equivalent
Thermoshaker for 2 mL tubes	Thermomixer® R with dry heating block 2 mL Eppendorf, catalog no. 022670107 and 022670549 or equivalent
Magnetic Separator	DynaMag™-15 magnet, Life Technologies, catalog no. 123.01D
Magnetic Separator	DynaMag™-2 magnet, Life Technologies, catalog no. 123.21D
Reference Pipettes with adjustable volumes of the following ranges 10-100 µL, 100-1000 µL	Eppendorf Research® Plus, adjustable-volume Pipette, Eppendorf, catalog no. 3120000020, 3120000046, 3120000062, or equivalent
Repeat Pipettor capable of repetitively dispensing volumes in an adjustable range	Eppendorf Repeater® Plus, Eppendorf, catalog no. 022260201, or HandyStep electronic, Brandtech®, catalog no. 705012, or equivalent
Bench Top Centrifuge with rotor for 1.5/2.0 mL tubes	Microcentrifuge 5430 R, Eppendorf, catalog no. 022620601 and 022636006, or equivalent
Multichannel Pipette	Eppendorf 8-Channel, 10-100 µL catalog no. 3122000035, or equivalent
Centrifuge for PCR plates	Centrifuge 4-16S, Qiagen Inc., catalog no. 81510 and 81031, or equivalent
100 mL graduated cylinder (PP)	Brandtech®, catalog no. V649941, or equivalent

General Laboratory Consumables and Reagents

Consumables and Reagents	Suggested Consumables and Reagents
Ethanol absolute, 200 proof, for molecular biology, ≥99.5 %	Sigma-Aldrich Co., catalog no. E7023, or equivalent
15 mL polypropylene (PP) centrifuge tubes with conical bottom, sterile	Sarstedt, catalog no. 62.554.205 PP, or equivalent
2.0 mL microtubes with round bottom and with attached PP cap with lid seal mechanism	Sarstedt SafeSeal, catalog no. 72.695.400, or equivalent
Pipette tips with aerosol barrier	Eppendorf ep Dualfilter T.I.P.S®: <ul style="list-style-type: none"> - 2-100 µL catalog no. 022491237, or equivalent - 50-1000 µL catalog no. 022491253, or equivalent
Repeat Pipettor Tips for volumes of 0.5 mL, 1 mL, 10 mL, 25 mL	Eppendorf Combitips Advanced® : <ul style="list-style-type: none"> - 0.5 mL catalog no. 0030089421, or equivalent - 1 mL catalog no. 0030089430, or equivalent - 10 mL catalog no. 0030089464, or equivalent - 25 mL catalog no. 0030089472, or equivalent
Disposable Transfer Pipettes, non-sterile bulk packaged, length about 15 cm (6 inches), stem diameter 5 mm, capacity about 5 mL	Samco™, Graduated 1 mL Large Bulb Non-sterile, Thermo Scientific catalog no. 222, or equivalent
Disposable Transfer Pipettes, non-sterile bulk packaged, length about 23 cm (9 inches), stem diameter 5 mm, capacity about 5 mL	Samco™ Extra Long Transfer Pipettes, Thermo Scientific, catalog no. 262, or equivalent
96 well plates for DNA storage	PCR Plate 96, skirted, Eppendorf, catalog no. 951020401, or equivalent
Adhesive Film or Foil for DNA storage plate	VWR® PCR Sealing Film catalog no. 82018-844, or equivalent
Applicator to form a tight seal between a microplate and an adhesive film	MicroAmp® Adhesive Film Applicator, Life Technologies, catalog no. 4333183, or equivalent
Cryo vials, 4 mL	VWR catalog no. 89094-812, or equivalent
Resealable bags, 10 x 15 cm, for disposing of used PCR plates	VWR catalog no. 89005-280, or equivalent

Required Special Equipment and Consumables

The following special equipment and consumables are needed to perform the Epi proColon test and cannot be replaced by other equipment.

Special Equipment/Consumables	Catalog and Part Numbers
Applied Biosystems® 7500 Fast Dx Real-Time PCR Instrument with Sequence Detection Software v1.4 21 CFR Part 11 Module and Windows XP operating system with Service Pack 2 or 3	Life Technologies Co., part no. 4406984 or 4406985
MicroAmp® Fast Optical 96-Well Reaction Plate with Barcode, 0.1 mL	Applied Biosystems (Life Technologies Co.), 20 plates, catalog no. 4346906, 200 plates, catalog no. 4366932
MicroAmp® 96- & 384-Well Optical Adhesive Film	Applied Biosystems (Life Technologies Co.) 25 sheets, catalog no 4360954 or 100 sheets, catalog no 4311971
BD Vacutainer®, K ₂ EDTA, 10 mL Blood Collection Tubes	Becton Dickinson, catalog no. 366643

Installation Requirements

The installation, calibration, performance verification and maintenance of the Applied Biosystems 7500 Fast Dx Real-Time PCR Instrument must be performed according to the manufacturer's instructions.

Note: Monthly background calibration as described in the manufacturer's maintenance procedure must be performed by the laboratory. Every 6 months the Operation Qualification and Performance Qualification Protocol (OQ/PQ) must be performed by an Applied Biosystems' Field Service Engineer. Semi-annual maintenance tasks are required for the Applied Biosystems 7500 Fast Dx PCR Instrument according to manufacturer's instructions, including calibration of the pure dyes FAM[™], JOE[™], TAMRA[™].

8. Specimen Collection and Handling

Blood Collection

Blood should be collected according to your laboratory's procedure for venipuncture using **ONLY** a **Vacutainer® K₂EDTA 10 mL blood collection tube** (purple top). Plasma preparation should be performed within 4 hours after blood is collected. The plasma sample may be stored refrigerated, at 2°C to 8°C for up to 72 hours, or frozen, at -15°C to -25°C for up to 14 days. Transport and shipment of refrigerated or frozen plasma should follow the same temperature and time requirements as stated above.

Note: Do not freeze **whole blood** samples.

Plasma Specimen Preparation

- Label all tubes with appropriate patient identification information
- Prior to use, disable the brake function in the centrifuge to prevent disruption of the cell layer
- Centrifuge the Vacutainer K₂EDTA blood collection tube for 12 min at 1350 ± 150 rcf. [For conversion of RPM (revolutions per minute) to rcf (relative centrifugal force), refer to the centrifuge manufacture's user manual]
- Remove blood collection tube from the centrifuge (If the plasma sample is hemolyzed, discard and acquire a new blood specimen from the patient)

- Using a fresh 6 inch disposable transfer pipette, transfer plasma from the blood collection tube to a 15 mL polypropylene centrifuge tube with conical bottom
- Centrifuge plasma in the 15 mL centrifuge tube for 12 min at 1350 ± 150 rcf
- Using a fresh, extra-long 9 inch disposable transfer pipette or serological pipette, transfer 3.5 mL of plasma into a labelled cryovial or centrifuge tube
- Start DNA extraction with the Epi proColon Plasma Quick Kit
- Alternatively, in a labelled cryovial, store plasma at 2°C to 8°C for up to 72 hours. For storage for up to 14 days, store the plasma at -15°C to -25°C.

Note: Take care not to disturb or transfer the buffy coat (white blood cells) layered above the red blood cells in the blood collection tube after the first centrifugation, or sedimented at the bottom of the conical centrifuge tube after the second centrifugation.

9. Test Procedure

A repeat pipettor is recommended for repetitive dispensing of the following reagents:

- Epi proColon Lysis-Binding Buffer
- Epi proColon Magnetic Bead Suspension
- Ethanol in DNA Binding Step 9.4
- Epi proColon Wash A Buffer
- Epi proColon Wash B Buffer
- Epi proColon Elution Buffer
- Epi proColon Bisulfite Solution
- Epi proColon Protection Buffer and the PCR Master Mix

Use of reference pipettes for pipetting extracted and bisulfite treated DNA is strongly recommended.

9.1. Preparation of Working Solutions

Preparation of Epi proColon Wash A Buffer

- Add 60.0 mL of Absolute Ethanol (for molecular biology, $\geq 99.5\%$) to the Epi proColon Wash A Concentrate using a sterile graduated cylinder or a serological pipette
- Close lid, mix thoroughly by inverting the bottle five times, avoiding formation of foam; Label the bottle with date of dilution, and mark the “Ethanol added” checkbox
- Store reconstituted Epi proColon Wash A Buffer at 15°C to 30°C for up to 6 weeks

Preparation of Epi proColon Wash B Buffer

- Add 40.0 mL of Absolute Ethanol (for molecular biology, $\geq 99.5\%$) to the Epi proColon Wash B Concentrate using a sterile graduated cylinder or a serological pipette
- Close lid, mix thoroughly by inverting the bottle five times. Label the bottle with date of dilution, and mark the “Ethanol added” checkbox
- Store reconstituted Epi proColon Wash B Buffer at 15°C to 30°C for up to 6 weeks

9.2. DNA Extraction and Bisulfite Conversion from Patient Plasma

The Epi proColon test kit contains sufficient reagents to process up to 32 samples including quality controls. One Epi proColon Positive Control and one Epi proColon Negative Control must be included in

each independent test run. There are 4 single-use tubes of Epi proColon Bisulfite Solution that will enable a maximum of 4 independent test runs (e.g., 4 runs of 8 samples each).

Note: Brief centrifugation of microtubes (stated as ‘Briefly spin down the tubes’) is required in several steps of this instruction to remove drops from the lid and/or to collect remaining liquid. It is recommended to centrifuge for 10 - 20 sec at $1,000 \pm 150$ rcf using a Bench-Top centrifuge. Avoid stronger centrifugation to prevent the compacting of magnetic bead pellets in these steps. For conversion of RPM (revolutions per minute) to rcf (relative centrifugal force), refer to the manufacturer’s centrifuge user manual.

Note: Vortexing of tubes and vials is required in several steps of this instruction to ensure homogeneous mixing of liquid. It is recommended to vortex samples for 5 to 10 seconds at medium speed.

Thawing of Plasma and Epi proColon Positive and Negative Control

- Thaw one Epi proColon Positive Control and one Epi proColon Negative Control for 30 min at 15°C to 30°C
- If frozen plasma samples are used, thaw samples for about 30 min at 15°C to 30°C
- Start Lysis within 60 min after thawing

9.3. Lysis

Note: Prior to use, briefly shake the Epi proColon Lysis Binding Buffer and visually check for precipitates. If precipitates are present heat the Epi proColon Lysis Binding Buffer in a water bath at 37°C for 60 min and shake gently until the precipitate is completely dissolved. Equilibrate Epi proColon Lysis Binding Buffer to room temperature before use.

- Add the following to a labelled 15 mL conical centrifuge tube:
 - 3.5 mL plasma sample, Epi proColon Positive Control, or Epi proColon Negative Control
 - 3.5 mL Epi proColon Lysis-Binding Buffer
- Cap the tube and mix by vortexing for 5 - 10 sec
- Incubate tubes on the bench top at 15°C to 30°C for 10 ± 1 min

9.4. DNA Binding

Note: A homogeneous suspension of the Epi proColon Magnetic Beads is essential for proper test performance. Deviations from the specified amount of beads may lead to false results. To ensure the correct magnetic bead concentration, the bottle should be mixed thoroughly just before pipetting. There should be no visible sediment at the bottom of the bottle. Mix to ensure a homogeneous suspension between the pipetting steps.

- Add to the 15 mL centrifuge tube in the following order:
 - 90 µL Epi proColon Magnetic Beads (freshly suspended)
 - 2.5 mL of Absolute Ethanol (for molecular biology, ≥ 99.5 %)
- Cap the tube and mix by inverting the tube 5-6 times
- Place 15 mL tubes into a rotator
- Rotate at room temperature for 45 ± 5 min at medium speed (approx. 10 - 20 rpm); adjust rotator angle to 35 – 45 degrees

9.5. DNA Washing

Note: Before starting the wash procedure set the thermoshaker to $80 \pm 2^{\circ}\text{C}$ for later use in the elution and bisulfite conversion steps.

- Place the 15 mL tubes into the DynaMag™-15 magnetic racks for 5 - 10 min
- Pour off the supernatant carefully, taking care not to remove magnetic beads
 - Add 1.5 mL Epi proColon Wash A Buffer
- Resuspend magnetic beads completely by vortexing for 5 - 10 sec
- Using an extra-long (9 inch) disposable transfer pipette, transfer magnetic beads suspension into a labelled 2.0 mL microtube
- Place disposable transfer pipette back into 15 mL tube for 2 - 6 min to collect remaining magnetic beads and transfer them into the 2.0 mL microtube
- Place the microtubes into the DynaMag-2 magnetic racks for 2 - 6 min
- Using a 6 inch disposable transfer pipette, remove as much buffer as possible while microtubes are still in the DynaMag-2 magnetic rack, taking care not to remove magnetic beads
- Briefly spin down the microtubes
- Place the 2.0 mL microtubes into the DynaMag-2 magnetic racks for 2 - 6 min
- Using a 10-100 μL reference pipette, remove as much residual buffer as possible while microtubes are still in magnetic rack

9.6. Elution

- Transfer the microtubes into a non-magnetic rack
- Vortex Epi proColon Elution Buffer for 5 - 10 sec
- Add 100 μL Epi proColon Elution Buffer to each microtube
- Close the microtubes
- Resuspend the magnetic beads by vortexing for 5 – 10 sec
- Place microtubes into a thermoshaker set to $1,000 \pm 100$ rpm and incubate at $80 \pm 2^{\circ}\text{C}$ for 10 ± 1 min
- Briefly spin down the microtubes
- Place microtubes into the DynaMag-2 magnetic racks for 2 - 6 min.
- Transfer the complete eluate, while microtubes are still in the magnetic rack, (~ 100 μL DNA solution) into fresh 2.0 mL microtubes
- Discard the 2 mL microtubes containing the magnetic beads

9.7. Storage of Extracted DNA

Note: If extracted DNA is not used immediately, store material at 2°C to 8°C for up to 24 hours. DO NOT FREEZE THE EXTRACTED DNA.

9.8. Bisulfite Conversion

Note: Epi proColon BISULFITE SOLUTION IS SENSITIVE TO OXYGEN CONTACT. Use only unopened tubes of Epi proColon Bisulfite Solution; do not store but discard any left-over solution.

- Add the following reagents to the 2.0 mL microtubes containing the eluate (~ 100 μL DNA solution):
 - 150 μL Epi proColon Bisulfite Solution

- 25 μ L Epi proColon Protection Buffer
- Cap the microtubes and mix the bisulfite reaction by vortexing for 5 - 10 sec
- Briefly spin down the microtubes
- Place microtubes into thermoshaker and incubate for 45 ± 5 min at $80 \pm 2^\circ\text{C}$ without shaking
- Remove microtubes from the thermoshaker immediately after 45 ± 5 min
- Reset thermoshaker temperature to $23 \pm 2^\circ\text{C}$, or set up a second thermoshaker to $23 \pm 2^\circ\text{C}$ for later use

9.9. Binding Step

Note: A homogeneous suspension of beads in the Epi proColon Magnetic Beads suspension is essential for proper performance. Deviations from the specified amount of beads may lead to false results. To ensure correct magnetic bead concentration, the bottle should be mixed thoroughly just before pipetting. There should be no visible sediment at the bottom of the bottle. Mix to ensure a homogeneous suspension between the pipetting steps.

- Briefly spin down the 2.0 mL microtubes containing the bisulfite reaction
- Add the following components to the microtube:
 - 1000 μ L Epi proColon Wash A Buffer
 - 20 μ L Epi proColon Magnetic Beads (freshly suspended)
- Mix by vortexing for 5 - 10 sec
- Wait until thermoshaker reaches $23 \pm 2^\circ\text{C}$
- Place the microtubes in the thermoshaker at 1000 ± 100 rpm and incubate for 45 ± 5 min at $23 \pm 2^\circ\text{C}$
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min.
- Using a fresh 6 inch disposable transfer pipette, remove as much liquid as possible while tubes are still in the magnetic rack, taking care not to remove magnetic beads

9.10. First Wash

- Remove the sample rack from the magnet for washing and vortexing
 - Add 800 μ L Epi proColon Wash A Buffer
- Resuspend by vortexing for 5 – 10 sec
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min.
- Using a fresh 6 inch disposable transfer pipette, remove as much liquid as possible while tubes are still in the magnetic rack, taking care not to remove magnetic beads

9.11. Second Wash

- Remove the sample rack from the magnet for washing and vortexing
 - Add 800 μ L Epi proColon Wash B Buffer
- Resuspend by vortexing for 5 – 10 sec
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min

- Using a fresh 6 inch disposable transfer pipette, remove as much liquid as possible while tubes are still in the magnetic rack, taking care not to remove magnetic beads

9.12. Third Wash

- Remove the sample rack from the DynaMag™-2 magnet for washing and vortexing
 - Add 400 µl Epi proColon Wash B Buffer
- Resuspend by vortexing for 5 – 10 sec
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min
- Using a fresh 6 inch disposable transfer pipette, remove as much liquid as possible while tubes are still in the magnetic rack, taking care not to remove magnetic beads
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min
- Using a 10-100 µL reference pipette, remove as much remaining liquid as possible while tubes are still in the magnetic rack, taking care not to remove magnetic beads

9.13. Drying

Note: Do not increase drying time or temperature as over-drying might reduce bisDNA recovery

- Open microtube lid
- Place open microtubes into thermoshaker
- Allow the pellet to dry for 10 ± 1 min at $23 \pm 2^\circ\text{C}$ without shaking

9.14. Elution

- Transfer microtubes into a non-magnetic rack
 - Add 60 µL Epi proColon Elution Buffer
- Close the microtubes
- Resuspend the magnetic beads by vortexing for 5 – 10 sec
- Incubate for 10 ± 1 min at $23 \pm 2^\circ\text{C}$ in a thermoshaker at 1000 ± 100 rpm
- Briefly spin down the microtubes
- Place microtubes on the DynaMag™-2 magnetic racks for 2 - 6 min
- Using a 10-100 µL reference pipette, transfer the complete eluate (~ 60 µL DNA solution) into a 96-well plate and seal the plate with adhesive film using an adhesive film applicator
- Set up the bisDNA storage plate according the recommended plate layout in Table 4

Table 4: Recommended Layout for a bisDNA Storage Plate.

	1	2	3	4	5	6	7	8	9	10	11	12
A	PC*	S7										
B	NC*	S8										
C	S1	S9										
D	S2	S10										
E	S3	S11										
F	S4	S12										
G	S5	S13										
H	S6	S14										

* PC: Epi proColon Positive Control, NC=Epi proColon Negative Control

9.15. Storage of bisDNA

If purified bisDNA is NOT USED IMMEDIATELY, store material at 2°C to 8°C for up to 24 hours or frozen at -25°C to -15°C for up to 3 days.

9.16. PCR Setup

Note: Each bisDNA sample (patient sample, Epi proColon Positive Control, or Epi proColon Negative Control) must be tested in triplicate.

Note: Prior to use, spin Epi proColon Polymerase for 10 - 20 sec at $1,000 \pm 150$ rcf using a Bench-Top centrifuge to remove drops from the lid.

Preparation of PCR Master Mix

- Thaw 1 or 2 Epi proColon PCR Mix tubes depending on the desired number of patient and control sample determinations (see Table 5)
- Vortex the Epi proColon PCR Mix tube(s) for 5 - 10 sec, briefly spin down the tube(s)
- Transfer the corresponding volumes of Epi proColon PCR Mix and Epi proColon Polymerase as indicated in Table 5 into a 2.0 mL microtube
- Mix the PCR Master Mix by vortexing for 5 - 10 sec
- Briefly spin the PCR Master Mix to remove drops from the lid

Note: Use PCR Master Mix, immediately. Do not store. Refreeze unused Epi proColon PCR Mix and Epi proColon Polymerase, directly after use.

Table 5: Preparation of PCR Master Mix.

Component	Volume for 8 Determinations (24 PCRs)	Volume for 16 Determinations (48 PCRs)	Volume for 24 Determinations (72 PCRs)	Volume for 32 Determinations (96 PCRs)
Epi proColon PCR Mix	383.8 µL	767.7 µL	1151.5 µL	1535.4 µL
Epi proColon Polymerase	19.4 µL	38.7 µL	58.1 µL	77.4 µL

Note: For a single PCR, 14.3 µL Epi proColon PCR Mix and 0.7 µL Epi proColon Polymerase are required.

PCR Plate Preparation

- Set up the PCR plate. The layout as shown in Table 6 is recommended
- Transfer 15 µl PCR Master Mix into the selected wells of the MicroAmp® Fast Optical 96-Well Reaction Plate
- Briefly centrifuge the bisDNA storage plate created in section 9.14 for 1 min at 1000 ± 100 rcf using the plate centrifuge
- Add 15 µl of each bisDNA solution to 3 independent wells of the PCR plate as shown in Table 6

Table 6: Recommended PCR Plate Layout

	1	2	3	4	5	6	7	8	9	10	11	12
A	PC*	PC*	PC*	S7	S7	S7						
B	NC*	NC*	NC*	S8	S8	S8						
C	S1	S1	S1	S9	S9	S9						
D	S2	S2	S2	S10	S10	S10						
E	S3	S3	S3	S11	S11	S11						
F	S4	S4	S4	S12	S12	S12						
G	S5	S5	S5	S13	S13	S13						
H	S6	S6	S6	S14	S14	S14						

*PC: Epi proColon Positive Control, NC=Epi proColon Negative Control

- Seal the plate with MicroAmp® Optical Adhesive Film
- Briefly spin down the plate with a plate centrifuge for 1 min at 1000 ± 100 rcf

Note: The filled PCR plate can be stored in a refrigerator at 2°C to 8°C for up to 4 h.

Plate Loading

Note: The PCR Master Mix does not contain ROX or any other reference dye. Accordingly, the passive reference setting must be set to “none”.

- Start the Software version SDS v1.4 21 CFR Part 11
- Create a new plate document
- Click “Create New Document”

- Define the following plate document:
 - Assay: Standard Curve (Absolute Quantification)
 - Container: 96-Well Clear
 - Template: Blank Document
 - Run Mode: Standard 7500
- Click “Next”
- Click “New Detector...”
- Create a new detector using following properties:
 - Name: Septin9
 - Description: Epi proColon
 - Reporter dye: FAM
 - Quencher dye: (none)
 - Color: Red
- Click “Create Another” and define following properties:
 - Name: ACTB
 - Description: Epi proColon
 - Reporter dye: JOE
 - Quencher dye: (none)
 - Color: Green
- Click “ok”
- Select both detectors and click “Add >>” to assign the detectors to the plate document
- Select “none” in the drop down menu of “Passive Reference”
- Click “Done”
- Go to the tab “Setup” and “Plate”
- Select all 96 wells of the plate
- Go to the menu point “View” and open the “Well Inspector”
- Select detector “Septin9” and “ACTB”
- Check the Passive Reference setting to be “none” (see Figure 1)
- Click “Close”
- Go to tab “Instrument” to program the cycling conditions as described in Table 7
- Change the following settings:
 - Sample Volume: 30 µl
 - Run Mode: Standard 7500
 - Data Collection: Stage 2, Step 2
- Create a “Thermal Profile” with 3 stages.
- Create a “Stage 2” having 3 steps, and a “Stage 1” and “Stage 3” having 1 step.
- Enter repetitions, target temperature, and hold time according to Table 7.

Change the “Ramp Rate” according to Table 7

- Set “Data Collection” for “Stage 2, Step 2 (55.5 @ 0:35)”
- Confirm the Thermal Cycler Protocol settings by comparing to Figure 2

- Save the run plate document under an appropriate file name
- A window will open asking for the “Reason for change of entry.” Enter “Setup” and any other comments relevant to the run
- Open the tray
- Place the PCR plate into the frame (position A1 goes to the upper left corner), ensure that the plate fits accurately in the frame. Close the tray.
- Start the run by pressing the “Start” button

Table 7: Thermal Cycler Program (Applied Biosystems 7500 Fast Dx with SDS v1.4)

Program Parameter	Denaturation	Cycling			Holding
Stage	"Stage 1"	"Stage 2"			"Stage 3"
Repetitions	1	45			1
Step	1	1	2	3	1
Target [°C]	94	62	55.5	93	40
Hold [mm:ss]	20:00	00:05	00:35	00:30	00:05
Auto Increment		0	0	0	
Ramp Rate [%]	40	80	80	40	80
Data Collection	Stage 2, Step 2 (55.5 @ 0:35)				

Figure 1: Screenshot from SDS v1.4 software after confirming settings in the 'Well Inspector' window.

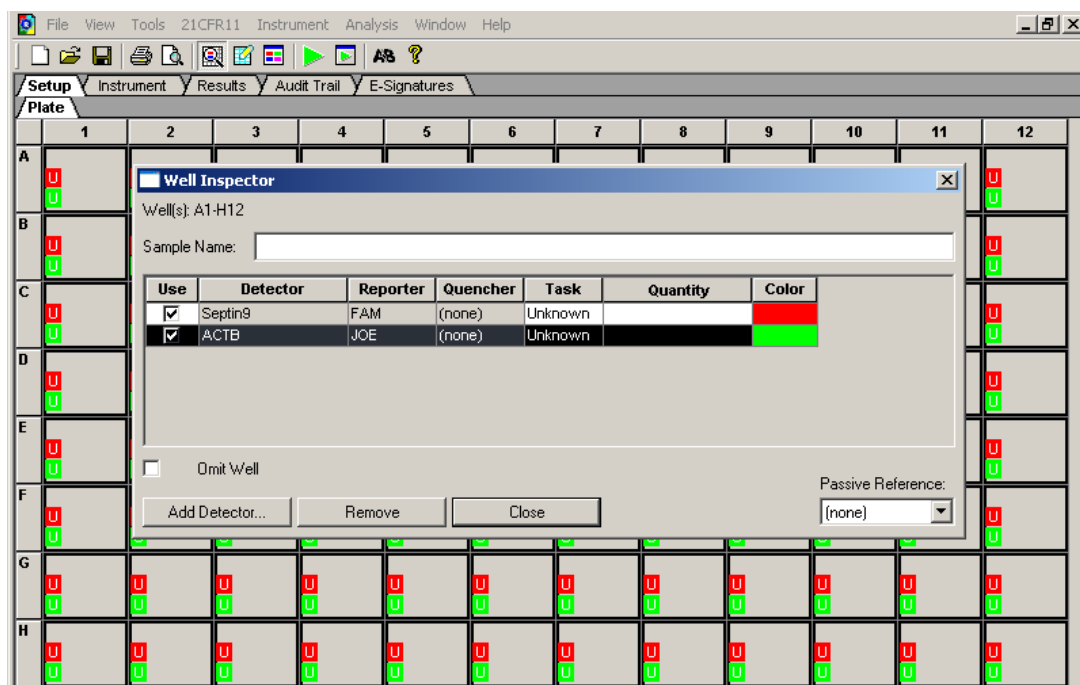
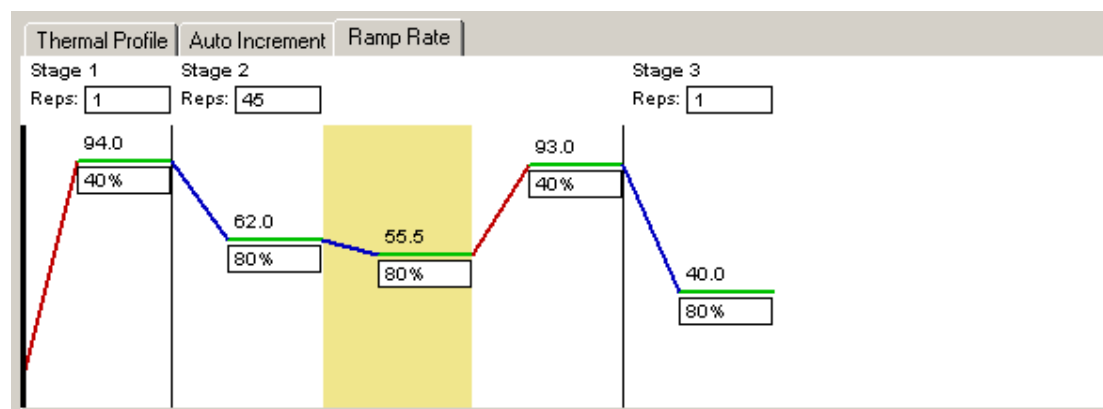
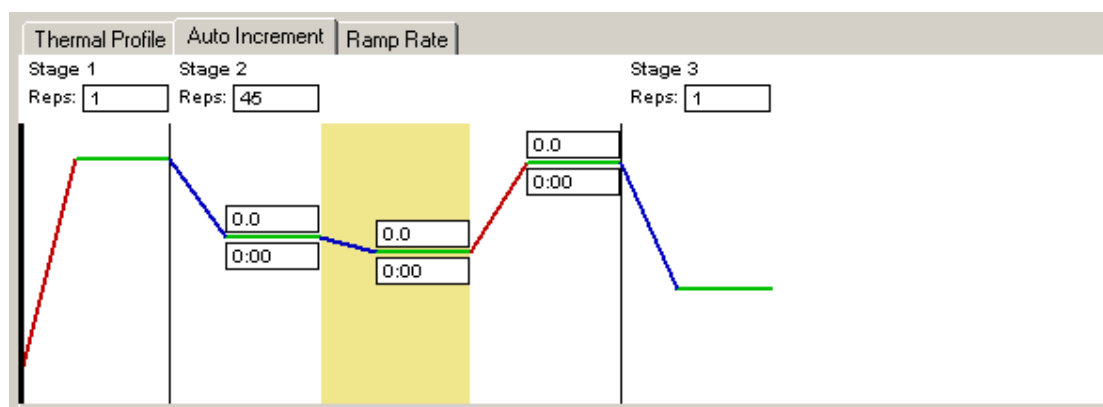
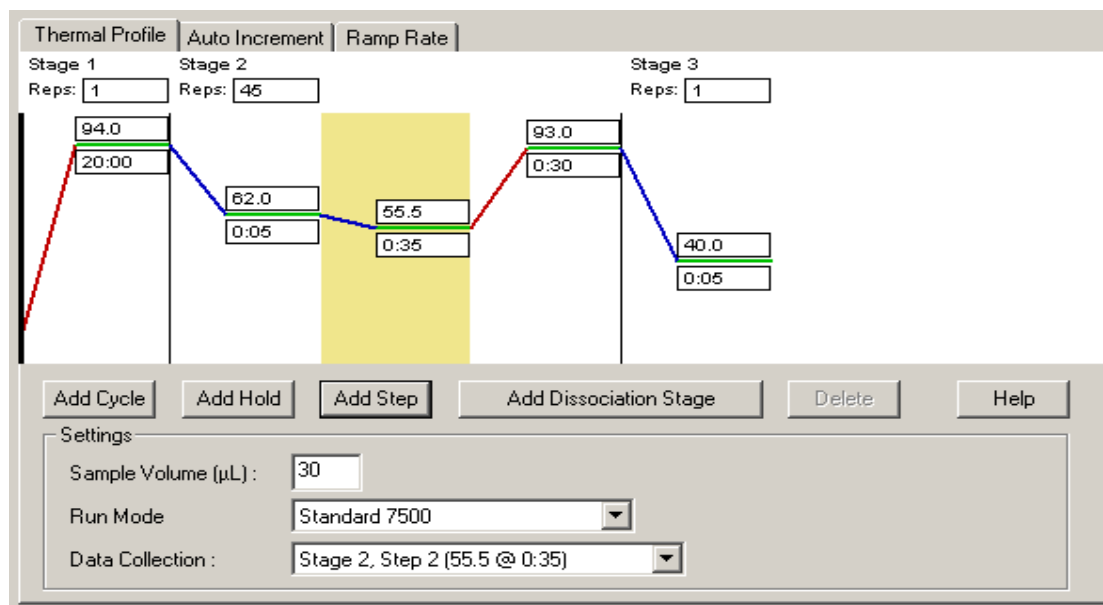



Figure 2: Screenshots from SDS v1.4 software showing correct settings for the cycling program. Temperature, time, repetitions should be displayed as shown in tab 'Thermal Profile' (upper). No changes of the default settings are made in the tab 'Auto Increment' (middle) and should be displayed as shown. Ramping rates should be displayed as shown in the tab 'Ramp rate' (lower). Sample volume, Run Mode and Data Collection are indicated below the cycling scheme in each of the tabs.



9.17. Results Analysis

Note: ANALYZE PCR RUNS WITH SOFTWARE VERSION SDS V1.4 ONLY. Software updates must not be initiated without written instruction from Epigenomics indicating that the Epi proColon test has been validated for use with the new software.

Note: Incomplete runs or runs where an error message occurs must not be analyzed. The run document must contain fluorescence data for 45 cycles.

- After completion of the PCR cycling program click “ok”
- Select the tab “Results”, then select the tab “Amplification Plot”
- Set “Analysis Setting” for Septin9 detector to be the following:
 - Data: “Delta Rn vs Cycle”
 - Detector: “Septin9”
 - Line color: “Detector Color”
 - Manual Ct, Threshold: “50000” (appears as “5.0e+004”)
 - Manual baseline, Start (cycle): “10”
 - Manual baseline, End (cycle): “22”
- Set “Analysis Setting” for ACTB detector to be the following:
 - Data: “Delta Rn vs Cycle”
 - Detector: “ACTB”
 - Line color: “Detector Color”
 - Manual Ct, Threshold: “25000” (appears as “2.5e+004”)
 - Manual baseline, Start (cycle): “10”
 - Manual baseline, End (cycle): “22”
- Confirm the correct Analysis Settings for each detector by comparing to Figure 3
- Analyze the data by pressing the “Analyze” button  in the top menu
- Save the file by pressing Save Document in the task bar. A window will open asking for the “Reason for change of entry”. Enter “Data analysis post run” and any other comments relevant to the run.
- Septin9 Ct values and ACTB Ct values are calculated automatically
- Ct values are displayed in the tab “Report”

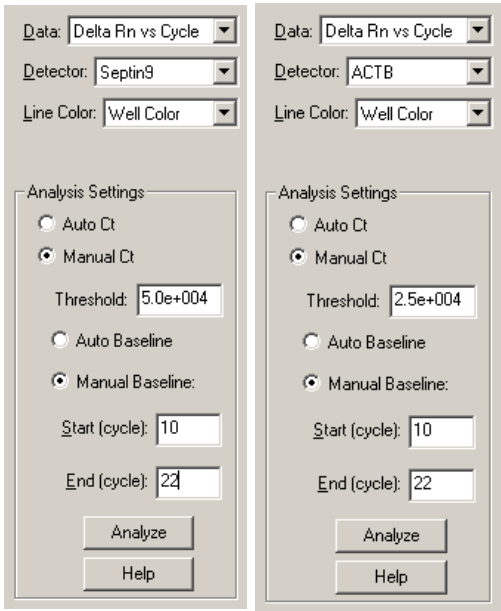


Figure 3: Screenshot from SDS v1.4 software showing correct analysis settings for the Septin9 detector (left) and ACTB detector (right).

10. Interpretation of Results

Assessment of the Validity of the Run by Epi proColon Controls

Any run of one or more patient sample(s) processed together with the Epi proColon Positive Control and Epi proColon Negative Control is considered VALID, when criteria set forth in Table 8 are met for **ALL THREE (3)** PCR replicates per control.

If either the Epi proColon Positive Control or Epi proColon Negative Control, or both is INVALID, the data for patient specimens processed together with the controls cannot be interpreted. Testing must be repeated for all patient specimens included in this test run.

Table 8: Validity Limits of Epi proColon Control Kit

Result of Control	Determination	Septin9 Result	ACTB Result
Positive Control VALID	PCR1	Ct* ≤ 41.1	Ct* ≤ 29.8
	PCR2	Ct* ≤ 41.1	Ct* ≤ 29.8
	PCR3	Ct* ≤ 41.1	Ct* ≤ 29.8
Negative Control VALID	PCR1	No Ct* result ("Undetermined")	Ct* ≤ 37.2
	PCR2		Ct* ≤ 37.2
	PCR3		Ct* ≤ 37.2

*cycle threshold

Assessment of the Validity of a Single PCR of a Patient Specimen

The interpretation of a single PCR is performed according to Table 9. Provided that the result of the internal control assay ACTB indicates sufficient input of bisDNA into the single PCR (ACTB cycle threshold at most 32.1), the result of the Septin9 PCR defines the result for this single PCR. An ACTB cycle threshold above 32.1 turns the single PCR result to "INVALID PCR".

Table 9: Interpretation of Results for Single PCR.

Single PCR Result	Septin9 Result	ACTB Result
Septin9 Positive	Ct* < 45	Ct* ≤ 32.1
Septin9 Negative	No Ct* result ("Undetermined")	Ct* ≤ 32.1
INVALID	Any result	Ct* > 32.1 Or "Undetermined"

*cycle threshold

Interpretation of Results for a Patient Specimen

The interpretation of patient test results is found on Table 10. The test result for a patient sample is "POSITIVE", if at least one PCR replicate is a Septin9 Positive PCR. The test result for a patient specimen is

“NEGATIVE”, if all three PCR replicates are Septin9 Negative PCRs. The test result is “INVALID” in all other cases.

Table 10: Interpretation of Epi proColon Test Results

Test Result	Positive Control Negative Control	Single PCR Results
POSITIVE	VALID	At least one Septin9 Positive PCR [†]
NEGATIVE	VALID	PCR1: Septin9 Negative PCR PCR2: Septin9 Negative PCR PCR3: Septin9 Negative PCR
INVALID	VALID	All other cases [‡]
INVALID	INVALID	n/a

[†] One single PCR result is Septin9 Positive; the two remaining single PCR results may have any result (INVALID, Septin9 Negative, or Septin9 Positive)

[‡] No single PCR result is Septin9 Positive and at least one single PCR result is INVALID; the remaining single PCR results may be INVALID or Septin9 Negative.

Note: The assessment of the validity and the interpretation of the Epi proColon test result are performed and recorded using the “The Epi proColon Results Form” available from the Epigenomics web site, www.epigenomics.com.

11. Quality Control

Quality Control procedures are designed to monitor the integrity of reagent performance and ensure the accuracy (validity) of reported test results. As with any control product or feature, patient results should not be reported if control results fail to yield the expected results. Each laboratory should refer to the guidelines and procedures established by their laboratory, appropriate regulatory agencies, or accrediting organizations in accordance to the requirements of each laboratory’s Quality Assurance program and accrediting organization.

The external Positive and Negative Controls are contained in the Epi proColon Control Kit (M5-02-003) for use with the Epi proColon Plasma Quick Kit (M5-02-001) and the Epi proColon PCR Kit (M5-02-002). The Internal Control (IC) is a part of the Epi proColon PCR Kit (M5-02-002).

External Controls: The Epi proColon Positive and Negative Controls kit (M5-02-003)

One Epi proColon Positive (PC) and one Negative (NC) Controls must be included with every run when testing patient samples using the Epi proColon test kit. These controls monitor the successful execution of the test workflow to ensure the validity of patient test results.

If the Epi proColon Positive and Negative Control test do not yield values within the required validity limits (see Table 8), see Remedial Actions, below.

Internal Control – Duplex β -Actin

For every PCR well, the duplexed detection of bisulfite-converted ACTB (β -actin) DNA serves as an internal control. This co-amplified sequence provides a monitor for sample quality, sample preparation and adequate DNA concentration of the specimen.

The Septin9 PCR test result is linked to the Ct value of the ACTB PCR (See Table 9). Ct values of the ACTB PCR outside the specified range INVALIDATE the single PCR test result; as such, high Ct values are associated with very low bisDNA or PCR inhibition.

If the Ct values of the ACTB reaction are outside the specified range and INVALIDATE the single PCR result, please refer to Remedial Actions, below.

Remedial Actions

When External Controls (PC, NC) fail to meet the validation criteria, patient test results for the entire run should be considered INVALID and should not be reported. Positive and Negative External Controls and patient samples should be repeated, and if INVALID results are obtained a second time, please contact Epigenomics Customer Support or your local distributor, for assistance.

For an Internal Control that does not meet validation criteria, that single PCR result is INVALID. The patient result should be assessed based on all 3 PCR results according to Table 10. If the patient result is INVALID based on this assessment, the result for the patient should not be reported.

Each laboratory should perform remedial actions according to their established Quality Assurance program prior to reporting patient test results.

12. Limitations of the Procedure

- This product has been validated for the combination of the Epi proColon Plasma Quick Kit (M5-02-001), the Epi proColon Sensitive PCR Kit (M5-02-002), and the Epi proColon Control Kit (M5-02-003) only. These kits and components (DNA extraction, bisulfite conversion or PCR kits) are not interchangeable or replaceable with other manufacturer's products.
- The Epi proColon test has been validated for use only with plasma derived from blood collected with BD Vacutainer® K₂ EDTA blood collection tubes (Becton Dickinson). Do not use this test with other clinical specimen types or with other blood collection tubes.
- The Epi proColon test has been validated for use only with the Applied Biosystems® 7500 Fast Dx Real-Time PCR Instruments with Sequence Detection Software v1.4 21 CFR Part 11 Module. Do not use with other instruments or software.
- Use of this test is limited to personnel experienced and trained in performing PCR assays. Good technique is essential and failure to follow instructions provided in these instructions may produce erroneous results.

13. Performance Characteristics**Clinical Sensitivity and Specificity**

Two studies were performed to demonstrate the clinical performance of the Epi proColon test. The Pivotal Clinical Study compared performance of the Epi proColon test to colonoscopy. In the

Supplemental Clinical Study, the performance of the Epi proColon test and a Fecal Immunochemical Test (FIT) were compared, using colonoscopy as the reference method.

For the Pivotal Study, plasma specimens from a study cohort (PRESEPT) were tested with Epi proColon at three independent clinical laboratories in the US. The PRESEPT clinical trial (ClinicalTrials.gov, Trial Registration ID: NCT00855348) enrolled 7,941 women (55%) and men (45%), average age of 60, from the CRC screening eligible population at average risk for CRC under the US Multi-Society Task Force Guideline at 32 clinical sites across the US and Germany.

In the Pivotal Study the Epi proColon test performance was determined from 1544 patients including all CRC patients, all patients with advanced adenomas (AA), and a random subset of subjects with small polyps (SP) or no evidence of disease (NED) from 6,857 eligible PRESEPT patients. Selection of SP and NED subjects was performed using a stratified random sampling approach based on the US 2010 census and the intended use population. Although not all the specimens in the PRESEPT study were tested, all relevant subpopulations were sampled and tested to derive the maximum value from the available sample pool. Clinical performance of the Epi proColon test was evaluated in terms of sensitivity for colorectal cancer and specificity in subjects negative for colorectal cancer, as determined by colonoscopy. The results for the Pivotal Study by overall clinical status (CRC and non-CRC) are shown in Table 11, with the sensitivity and specificity outcome in Table 12.

Table 11: Result of Epi proColon Testing Sample Type

Sample Status	Negative	Positive	VALID Samples
CRC	14	30	44
Non-CRC	1182	318	1500
Total	1196	348	1544

Table 12: Clinical Performance of Epi proColon (Pivotal Study)

Sample Status	Point Estimate	Lower 95%	Upper 95%
Sensitivity	68.2 %	53.4 %	80.0 %
Specificity	78.8 %	76.7 %	80.8 %
Specificity* (weighted)	79.1 %	77.0 %	81.4 %
Specificity** (weighted)	80.0%	77.9 %	82.1 %

* weighting according to US census population

** weighting according to PRESEPT patient disposition

In the Supplemental Study the clinical performance of the Epi proColon test was compared to the OC FIT-CHEK® (Polymedco, Inc.) test in a multi-center study. The study was designed to collect matched blood and fecal specimens and clinical data from screening guideline-eligible subjects using colonoscopy as the reference method.

Subjects were recruited at 61 clinical sites in the US according to the following scheme:

- Subjects having CRC or a high suspicion of invasive CRC identified during screening colonoscopy were enrolled and provided blood and fecal samples at least 10 days after colonoscopy but prior to surgery or intervention
- Prospectively enrolled subjects provided blood and fecal samples prior to bowel prep for screening colonoscopy

Of 337 subjects enrolled in the study, 36 were excluded due to failure to meet inclusion/exclusion criteria. From the remaining 301 enrolled subjects, there were 101 CRC, 29 AA, 77 SP and 94 NED. Plasma samples were available from all 301 subjects. Fecal samples were not available from 11 subjects (4 CRC, 2 AA, 2 SP and 3 NED).

Both, the Epi proColon testing and OC FIT-CHEK testing were performed at an independent US diagnostic laboratory. Test outcome was compared to results obtained from colonoscopy for both methods.

Overall sensitivity and specificity for the Epi proColon test and FIT test for all available samples and on paired samples as compared to the reference colonoscopy are calculated in Tables 13a, b and c, with a three-way summary in Table 14.

The observed sensitivity for CRC on paired samples was 4.2% higher for the Epi proColon test. The 95% confidence interval (-16.2%; 8.1%) was strictly below the non-inferiority margin of 10% pre-set in the protocol. The sensitivity of the Epi proColon test is statistically non-inferior to the FIT test.

For specificity, the difference between tests was 16.6% in favor of the FIT with a 95% confidence limit (10.6%; 22.9%) around the estimate. This result does not demonstrate non-inferiority for specificity when compared to the non-inferiority margin of 20% pre-set in the protocol.

Table 13a: Epi proColon Sensitivity and Specificity, n = 301 All Plasma Samples

Parameter	Point Estimate	CI 95%
Sensitivity	73.3% (74/101)	63.9 – 80.9%
Specificity	81.5% (163/200)	75.5 – 86.3%

Table 13b: Epi proColon Sensitivity and Specificity, n = 290 Paired Samples

Parameter	Point Estimate	CI 95%
Sensitivity	72.2% (70/97)	62.5 – 80.1%
Specificity	80.8% (156/193)	74.7 – 85.8%

Table 13c: FIT Sensitivity and Specificity, n = 290 Paired Samples (equals number of available fecal samples)

Parameter	Point Estimate	CI 95%
Sensitivity	68.0% (66/97)	58.2 – 76.5%
Specificity	97.4% (188/193)	94.1 – 98.9%

Table 14: Three-Way Comparison of the Epi proColon and FIT Tests, and Colonoscopy (paired samples)

Diagnostic Accuracy Criteria: Standard Colonoscopy						
Colorectal Cancer				Non-Colorectal Cancer AA, SP, NED		
	Epi proColon Positive	Epi proColon Negative	Total	Epi proColon Positive	Epi proColon Negative	Total
FIT Positive	50	16	66	1	4	5
FIT Negative	20	11	31	36	152	188
Total	70	27	97	37	156	193

A statistical comparison for the rate of cancer detection (sensitivity) according to the study protocol demonstrated that the Epi proColon test was not inferior to the FIT test.

Analytical Sensitivity - Limit of Detection

The Limit of detection (LoD) of the Epi proColon test was determined at four laboratories according to CLSI EP12 and EP17 guidance documents. All sites utilized the test kit manufactured under final manufacturing conditions. Seven levels of technical samples were tested with a range of methylated DNA from 0 (blank) to 50 pg/mL. LoDs were determined using DNA spiked into plasma and into an artificial matrix of Tris buffer plus BSA. Based on the results of this study, the LoD is 4.7 pg/mL (95% CI 2.5-9.0 pg/mL).

Precision and Accuracy

Accuracy – Internal study

The assay Ct thresholds for the Epi proColon test were established in a study performed using 156 clinical plasma samples (reference method: colonoscopy). The instrumentation was set to 50 cycles in order to establish the Septin9 and ACTB cycle threshold (Ct) limits on these clinical specimens. From the distributions of these data, it was evident that Ct values larger than 40 for Septin9 PCR are rare and no Ct-values are observed above a Ct of 45. The vast majority of ACTB measurements range between 26 to 30 Ct. There was a general observation in these clinical specimens of outliers having lower Ct-values. For very few specimens the ACTB value came close to Ct of 31. From these data Septin9 and ACTB Ct limits of 45 and 32.1, respectively, were determined.

Accuracy – Cut-off Verification

A second study was performed using 346 clinical plasma samples. The instrumentation was set at 50 cycles for a subset of 197 plasma samples in order to verify the Septin9 and ACTB cycle threshold limits of 45 and 32.1, respectively, on these clinical specimens. The distributions of these data confirmed the results from the first study. Therefore, the cycle threshold limits were verified. For these 346 clinical samples, the overall agreement with clinical status was observed at 94.9% (93/98) with a 95% CI from 88.6% – 97.8% for CRC cases, and 84.3% (209/248) with a 95% CI from 79.2% – 88.3% for colonoscopy-verified controls.

Repeatability, Intermediate Precision and Reproducibility

To evaluate repeatability and intermediate precision of the assay, aliquots from 14 clinical sample pools were tested at three testing sites by six operators with three reagent lots using three PCR instruments. Each plasma pool was tested 12 times. Pools 1 – 6 generated from CRC plasma were positive in all 12 tests. For the three pools representing self-declared healthy blood donors, pool 7, pools 8 and pool 9 were each negative in 9 out of 12 tests. For the pools derived by diluting a single CRC plasma aliquot in human plasma, pools 10, 13, and 14 were positive in 11 out of 12 tests, while pools 11 and 12 were positive in all 12 replicates. In total, for 129 out of 132 samples where CRC plasma was tested (pools 1 – 6, pools 10 – 14), the test result was positive leading to 98% (95% CI: 94 – 99%) positive percent agreement with clinical status. Aggregated over the three pools derived from healthy blood donors (pools 7 – 9), the test results for 27 out of 36 samples was negative leading to 75% (95% CI: 59 – 86%) negative percent agreement with clinical status. The total percent agreement estimated from these data is 156/168, i.e. 93% (95% CI: 88 – 96%). There were no differences in the positive and negative percent agreement attributable to sites, operators or kit lots.

In addition, precision and reproducibility analyses were conducted with Ct values generated on the set of 14 sample pools. For precision, the ranges of standard deviation and coefficient of variation (CV) for Septin9 were 0.4 – 2.1 Ct and 1.1 – 5.5%, respectively. The corresponding ranges for ACTB were 0.2 – 0.4 Ct and 0.8 – 1.7%.

For reproducibility, the ranges of standard deviation and coefficient of variation for Septin9 were 0.4 – 2.3 Ct and 1.4 – 6.0%, respectively. The corresponding ranges for ACTB were 0.2 – 0.4 Ct and 0.7 – 1.6%.

Cross-Reactivity

To test for sequence cross-reactivity, BLAST alignment searches and electronic PCR analyses were performed against the human genome with the Septin9 PCR assay (blocker, primers and probe). This analysis showed that the test is specific for amplification of the bisDNA sequence of methylated Septin9 and therefore showed no cross-reactivity within the human genome.

364 samples from patients with other cancers (prostate, breast, lung) or other diseases (hypertension, hyperlipidemia, diverticulitis, chronic gastritis, or cardiovascular) with no evidence of CRC by colonoscopy, were evaluated to determine cross-reactivity. The Epi proColon test was positive in 72 of 173 patients (42%) with other cancers and positive in 33 of 191 patients (17%) with other diseases. The methylation status of the Septin9 region was determined for 7 positive patients (3 patients with other cancers, 4 patients with other diseases) by bisulfite sequencing of the PCR product. For all test positives, sequencing confirmed methylation of the Septin9 PCR target area. This study demonstrates that the Epi proColon test detects the true biological status for these other conditions rather than reporting cross-reactivity.

Interfering Substances

A study was conducted to verify that the presence of interfering substances potentially found in plasma samples have no effect on the test results. The ten (10) most common substances present in human plasma were selected and tested at the highest concentration that would occur in a clinical setting. Albumin (40 mg/mL), Bilirubin (0.2 mg/mL), Cholesterol (5 mg/mL), D-(+) Glucose (10 mg/mL), Hemoglobin (10 mg/mL), K₂EDTA (20 mg/mL), Red Blood Cells (0.4 % v/v), Triglycerides (12 mg/mL), human genomic DNA (100 ng/mL), and Uric Acid (0.235 mg/mL) were tested in 6 plasma specimen each (3 analyte negative plasma specimen, 3 analyte positive plasma specimen). None of the substances interfere with the test when added to the specimen at the listed concentration level.

14. Meaning of Symbols

Consult Instructions for Use



Order Number

*In Vitro* Diagnostic Medical Device

Lot Number



Expiration Date



Manufacturer



Store at indicated Temperature



Number of Tests



Do not Reuse

15. References

- 1 deVos, T et al. Circulating methylated SEPT9 DNA in plasma is a biomarker for colorectal cancer. *Clinical Chemistry* 55:7, 1337-46 (2009)
- 2 Lofton-Day, C. et al. DNA methylation biomarkers for blood-based colorectal cancer screening. *Clinical Chemistry* 54:2, 414-423 (2008)
- 3 Gruetzmann, R. et al. Sensitive Detection of Colorectal Cancer in Peripheral Blood by Septin 9 DNA Methylation Assay. *PLoS ONE*, Volume 3, Issue 11, e3759 (2008)
- 4 Model, F. et al. Identification and Validation of colorectal neoplasia-specific methylation markers for accurate classification of disease. *Mol Cancer Res* 5, 153-163 (2007)
- 5 Eads, C.A. et al. MethyLight: a high-throughput assay to measure DNA methylation. *Nucleic Acids Research*, Volume 28, Issue 8, e32 (2000)
- 6 Warren, D. et al. Septin 9 methylated DNA is a sensitive and specific blood test for colorectal cancer. *BMC Medicine*, 9:133 (2011)
- 7 Epigenomics data on file.

16. Contact Information

For questions, information or Customer Support, please contact Epigenomics in one of the following ways:

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